Synthesis of Oxime-Linked Mucin Mimics containing the Tumor-Related T_N and Sialyl T_N Antigens

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ABSTRACT

The synthesis of oxime-linked mucin mimics was accomplished via the incorporation of multiple ketone residues into a peptide followed by reaction with aminooxy sugars corresponding to the tumor-related T_N and sially T_N (ST_N) antigens.

The site-specific attachment of oligosaccharides to proteins can be accomplished by reaction of nucleophilic sugar derivatives with aldehydes and ketones.¹ Previously, we reported on the use of ketone-amino acid 1 (Figure 1) for the synthesis of glycopeptide analogs containing unnatural sugar-peptide linkages.² Amino acid 1 can be prepared in one step via reductive ozonolysis of commercially available Fmoc-dehydroleucine (2)³ and can be incorporated into peptides (3) by Fmoc-based solid-phase peptide synthesis (SPPS) without need for protection

of the ketone group.⁴ The ketone group is chemically orthogonal to all naturally occurring amino acid side chain functional groups and thus can be selectively condensed with aminooxy sugars to give the corresponding oximelinked products (4). Since glycoproteins often contain more than one site of glycosylation, we were interested to see if this strategy could be applied to the synthesis of glycopeptides with clustered oxime-linked glycans.

Mucins are a class of heavily *O*-glycosylated proteins that are abundantly secreted by epthelial cells specialized for mucus production.⁵ They are rich in serine and threonine residues bearing α-linked glycans initiated by *N*-acetylgalactosamine (GalNAc). A variety of carbohydrate ligands required for cell-surface interactions, including the Lewis and blood group antigens, can be presented on a

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mucin scaffold. In cancer cells, the glycosylation pattern of mucins is altered, leading to the expression of distinct tumor-related epitopes. Thus, mucin fragments and their cancer-associated oligosaccharides have attracted much attention as components of synthetic cancer vaccines. In the present study we focused on the preparation of oximelinked mucin mimics containing clusters of the T_N and sialyl T_N (ST_N) antigens (Figure 2), which are abundantly expressed in many types of cancer, including tumors of the breast, colon, liver and pancreas.

Figure 1. Synthesis of oxime-linked glycopeptides.

We chose fragments of the endothelial mucin GlyCAM-1 (5 and 6, Figure 3) as peptide scaffolds. Peptides 5 and 6 each replace six Ser or Thr residues within the 12- or 17-amino acid sequence with amino acid 1 (designated with the single letter code "O"). The syntheses of 5 and 6 were carried out on an automated peptide synthesizer using DCC/HOBt mediated couplings, on MBHA and Fmoc-Glu(tBu)-Wang resins, respectively. Amino acid 1 was used without protection of the ketone group. Following chain assembly the peptides were cleaved from the resin

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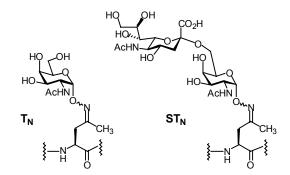


Figure 2. Oxime-linked analogs of the T_N and ST_N antigens.

GlyCAM-1:

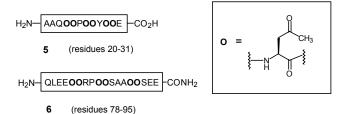


Figure 3. Amino acid sequence of GlyCAM-1 and corresponding peptide fragments 5 and 6 bearing ketone-amino acid 1 (single letter code "O") in place of Ser and Thr.

The synthesis of the aminooxy T_N antigen (7) was accomplished as previously described using a phase transfer catalyzed (PTC)¹² reaction of *N*-hydroxysuccinimide (NHS) with glycosyl chloride 8^{13} (Scheme 1). Reductive acetylation of 9, followed by mild hydrazinolysis afforded the desired aminooxy sugar (7). We have recently found that the intermediate NHS

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 $^{^{11}}$ ESI-MS (neg-ion mode): calcd for **5** 1356.1, found 1355.7; calcd for **6** 2021.8, found 2021.7.

glycoside (9) can also be accessed using glycosyl bromide 10^{13} as a donor in a Koenigs-Knorr glycosylation with NHS. While the stereoselectivity of this reaction is not as high as that achieved in the PTC reaction, the preparation of the α -bromide is generally higher yielding than the β -chloride (95% versus 45%)¹³ making this route an attractive alternative.

Scheme 1^a

^aReagents: (a) NHS, (nBu)₄NHSO₄, CH₂Cl₂, Na₂CO₃, 57% (α only); (b) NHS, AgClO₄, CH₂Cl₂, 4 Å MS, 67% (3:1 α/β); (c) H₂, Pd/C, Ac₂O, 100%; (d) 10% aq. N₂H₄, 71%.

The synthesis of aminooxy-ST_N 11 utilized the selectively-protected glycosyl acceptor (12), containing the pre-installed NHS glycoside, for reaction with known sialyl phosphite 13¹⁴ (Figure 4). For the installation of the NHS glycoside we chose to use a Koenigs-Knorr glycosylation with glycosyl bromide 14, which was obtained from 6-*O*-TBDPS-D-galactal (15).¹⁵

As depicted in Scheme 2, glycosyl bromide **14** was generated in three steps via iosopropylidine formation, azidonitration with CAN and NaN₃ and treatment with LiBr. Reaction of bromide **14** with NHS in the presence of AgClO₄ gave compound **16** in 65% yield as a mixture of anomers (3:1 α/β). Isolation of the desired α -glycoside (**12**) was achieved following removal of the TBDPS group with TBAF. Glycosylation of **12** with sialyl phosphite **13** using TMSOTf as the promoter gave disaccharide **17** in 44% yield as a mixture of anomers (3:1 α/β). Subsequent deprotection and reductive acetylation of **17** over a series of steps afforded the target aminooxy-ST_N **11**.

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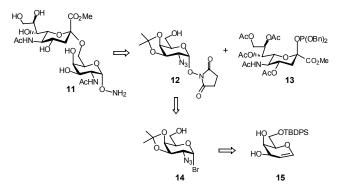
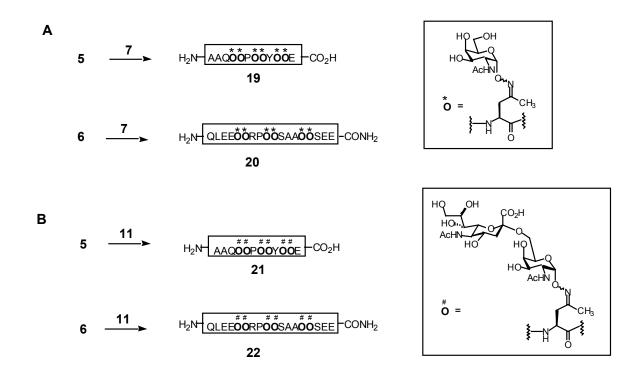


Figure 4. Retrosynthesis of aminooxy-ST_N (11).

Scheme 2^a

^aReagents: (a) Me₂C(OMe)₂, PPTS, DMF, 50 °C, 1 h, 95%; (b) CAN, NaN₃, CH₃CN, -20 °C, 15 h, 71%, (c) LiBr, CH₃CN, rt, 5 h, 96%; (d) NHS, AgClO₄, CH₂Cl₂, 4 Å MS, rt, 2 d, 65% (3:1 α/β); (e) TBAF, AcOH, THF, rt, 6 h, 45%; (f) **13**, TMSOTf, THF, 4 Å MS, -35 °C, 1 h, 44% (3:1 α/β); (g) *p*-TsOH, MeOH, rt, o.n., 60%; (h) Ac₂O, pyridine, DMAP, rt, o.n., 54%; (i) H₂, Pd/C, Ac₂O, rt, 2 h, 49% after HPLC; (j) i) NaOMe, MeOH, rt, 24 h, ii) LiOH, MeOH, H₂O, 4 °C, o.n., iii) 10% aq. N₂H₄·H₂O, 70% for 3 steps.

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Scheme 3. Synthesis of mucin mimics 19 and 20, containing the T_N antigen (A) and mucin mimics 21 and 22, containing the ST_N antigen (B). All ligation reactions were performed by incubating the peptide with an excess of aminooxy sugar at 37 °C in NaOAc buffer, pH 5.5.

The ligation of 7 and 11 with peptides 5 and 6 was carried out at 37 °C with an excess of either sugar in NaOAc buffer, pH 5.5 (Scheme 3). The reactions were monitored by reversed-phase HPLC and judged to be complete after 24 h. The oxime-linked products (19-22) containing the $T_{\rm N}$ and $ST_{\rm N}$ antigens were purified by reversed-phase HPLC (60-70% yield) and their identity confirmed by ESI-MS. ¹⁶

These syntheses illustrate that multiple clustered ketone residues can be incorporated into a peptide and reacted with aminooxy sugars. Such an approach circumvents the need to synthesize large quantities of complex glycosyl amino acids for use in peptide synthesis, a process which can be extremely labor intensive depending on the complexity of the pendant glycan. The oxime-based strategy benefits from convergent assembly of peptides and aminooxy sugars, both of which are straightforward to prepare. The incorporation of these glycopeptides into larger, full-length proteins, by techniques such as native and expressed protein ligation, ¹⁷

should provide access to homogenous mucin-analogs for a variety of applications.

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Supporting Information Available: Full experimental procedures and tabulated ¹H and ¹³C NMR data for all compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

¹⁶ ESI-MS (neg-ion mode): calcd for **19** 2665.3, found 2665.6; calcd for **20** 3331.3, found 3330.2; calcd for **21** 4412.9, found 4413.2; calcd. for **22** 5078.9, found 5080.0.

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